

Plant Archives

Journal homepage: http://www.plantarchives.org
DOI Url: https://doi.org/10.51470/PLANTARCHIVES.2024.v24.no.2.222

EXPLORING MULTI-FACETED APPROACHES IN COTTON BREEDING FOR PINK BOLLWORM RESISTANCE

J.D. Patel^{1*}, Nidhi Dubey¹, I.R. Delvadiya¹, S. Anvesh¹, Iram Bashir² and Rajneesh Kumar²

¹Department of Genetics and Plant Breeding, School of Agriculture, Lovely Professional University, Phagwara - 144 411 (Punjab), India.

²Division of Genetics and Plant Breeding, Faculty of Agriculture (FoA), SKUAST–K, Wadura - 193 201, J & K, India. *Corresponding author E-mail: jaydeeppatel152@gmail.com (Date of Receiving-22-06-2024; Date of Acceptance-28-08-2024)

(

ABSTRACT

The pink bollworm (Pectinophora gossypiella) poses a significant threat to global cotton cultivation, causing substantial economic losses and environmental harm. The ramifications of pink bollworm infestations are severe, imposing significant economic burdens on cotton producers. Immediate financial losses occur as yields plummet and cotton quality deteriorates. The burden increases with the need for pest management strategies, requiring additional investments in resources and labor. Traditional pest management methods, reliant on chemical pesticides, exacerbate ecological imbalances and contribute to pesticide resistance. Sustainable alternatives, such as integrated pest management (IPM), offer comprehensive solutions by combining biological, cultural and chemical interventions tailored to specific contexts. However, the emergence of pest resistance necessitates ongoing innovation in pest management techniques. Emerging technologies, including precision agriculture, remote sensing and genetic engineering, hold promise for revolutionizing pest management practices. These advancements enable targeted application of inputs, early pest detection, and the development of resistant cotton varieties. Furthermore, multi-omics approaches and genome editing technologies provide insights into the molecular mechanisms underlying pest resistance, facilitating the development of resilient cotton cultivars. The future of cotton breeding for sustainable pest management lies in the integration of these technologies, ensuring the long-term viability of cotton agriculture while minimizing environmental impact.

Key words: Pink bollworm, Cotton, Pest management, Resistance breeding, Biological control.

Introduction

The pink bollworm (*Pectinophora gossypiella*) poses a severe threat to global cotton cultivation, having spread from its origins in Asia to infest cotton fields worldwide. This pest completes its life cycle within the cotton plant, particularly targeting the bolls. The larvae burrow into the bolls, consuming seeds and shredding fibers, leading to significant declines in both the quantity and quality of cotton lint produced (Tabashnik *et al.*, 2022). Their damage results in substantial economic losses for cotton producers, necessitating increased investments in pest management strategies (Smith *et al.*, 2019).

The ramifications of pink bollworm infestations are severe, imposing significant economic burdens on cotton producers. Immediate financial losses occur as yields plummet and cotton quality deteriorates. The burden increases with the need for pest management strategies, requiring additional investments in resources and labor (Vennila *et al.*, 2021). Resorting to chemical pesticides presents further ecological risks, as these substances harm non-target species, contaminate soil and water, and promote pesticide-resistant insect populations. These effects extend beyond agricultural ecosystems, disrupting broader ecological balances (Sharma *et al.*, 2022). The reliance on chemical pesticides fosters a vicious cycle, with pest resistance necessitating increasingly potent

agents, escalating environmental harm and imposing economic burdens due to rising costs and diminishing returns. Sustainable alternatives, such as integrated pest management (IPM), offer comprehensive solutions by combining biological, cultural, and chemical interventions tailored to specific contexts (Gore *et al.*, 2022). IPM promotes natural predators, pheromone traps, and pestresistant cotton varieties to mitigate pest pressures while minimizing environmental impact and preserving ecosystem integrity. This approach underscores the need to balance agricultural productivity with ecological stewardship (Dhillon *et al.*, 2021).

Developing effective strategies for pest resistance in cotton necessitates a thorough understanding of the underlying molecular mechanisms. Multi-omics approaches, encompassing genomics, transcriptomics, proteomics and metabolomics, provide comprehensive insights into the complex biological interactions within plants under pest stress. These methods have identified key regulatory networks and metabolic pathways that can be manipulated to enhance resistance (Razzaq et al., 2019). High-throughput sequencing technologies have significantly advanced our ability to assemble high-quality cotton genomes, revealing crucial structural variations and genetic markers associated with resistance traits (Peng et al., 2020). The integration of multi-omics data with traditional and molecular breeding techniques heralds a new era of precision breeding. This holistic approach not only improves pest resistance, such as against the pink bollworm, but also supports sustainable agricultural practices by reducing dependency on chemical pesticides and promoting environmental health (Vaughan et al., 2018). These methodologies are increasingly critical as climate change exacerbates challenges in cotton production, necessitating the development of resilient varieties.

Traditional breeding methods: A foundation for resistance

Overview of selective Breeding

Selective breeding, or artificial selection, has been a fundamental practice in agricultural development since the dawn of farming. In cotton breeding for pest resistance, selective breeding involves the intentional selection of parent plants exhibiting desirable traits to produce offspring that inherit these characteristics. The objective is to amalgamate multiple advantageous traits, such as high yield, quality fiber, and pest resistance, into new cotton varieties capable of thriving under various environmental conditions and resisting pest pressures, notably those posed by the pink bollworm (*Pectinophora*

gossypiella). The economic and environmental consequences of pink bollworm infestation in cotton agriculture tabulated in Table 1.

Steps in the selective Breeding Process Germplasm collection and Evaluation

The initial phase of selective breeding involves the collection and evaluation of a diverse range of cotton germplasms. These germplasms include both wild relatives and cultivated varieties that exhibit a spectrum of traits, including potential resistance to pests. Researchers conduct controlled infestation trials and field evaluations to assess these germplasms for inherent resistance traits. This evaluation is crucial for identifying promising genetic reservoirs that can be utilized in breeding programs (Brar *et al.*, 2018).

Progeny selection and Evaluation

The progeny resulting from controlled crosses undergo extensive selection protocols. Researchers employ a combination of phenotypic assessments and genetic analyses to identify individuals exhibiting the desired resistance traits along with optimal agronomic performance. This step often involves multi-year field trials to evaluate the stability and effectiveness of the resistance traits under various environmental conditions, ensuring that selected lines can perform consistently across different settings (Liu *et al.*, 2017).

Backcrossing and Line Development

To ensure that the new varieties retain resistance traits while enhancing other characteristics, breeders may employ backcrossing. This process involves repeatedly crossing the progeny back to one of the parent varieties to stabilize the desired traits within the population. Over several generations, breeders develop stable lines that consistently express both the desired resistance and superior agronomic traits (Smith *et al.*, 2012).

Varietal Refinement and Testing

The final step in the selective breeding process involves the refinement and rigorous testing of superior lines. This phase includes extensive field trials across different regions and environmental conditions to ensure that the new varieties are adaptable and perform well under diverse conditions. The objective is to maximize the durability and performance of the new cultivars before they are released to farmers, thereby ensuring their commercial viability and sustainability (Tabashnik *et al.*, 2013).

In cotton breeding, a thorough understanding of the pink bollworm (*Pectinophora gossypiella*) life cycle is critical for developing resistant varieties. Breeding

Table 1: The economic and environmental consequences of pink bollworm infestation in cotton agriculture.

Consequences	Economic Properties	Environmental Properties	
Crop Losses	Reduced yields (Pimentel et al., 1997)	Decreased biodiversity (Tilman et al., 2006)	
Revenue Reduction	Lower marketable cotton production (Altieri et al., 2003)	Impact on natural predators and beneficial insects (Gurr et al., 2017)	
Increased Costs	Higher input costs for pest control (Pretty, 2007)	Soil degradation (Lal, 2009)	
Market Instability	Price fluctuations due to supply shortages (Godfray et al., 2010)	Ecosystem disruption (Chapin III et al., 2000)	
Loss of Livelihoods	Impacts on farm incomes and rural economies (Pretty et al., 2000)	Disruption of food webs (Tylianakis <i>et al.</i> , 2008)	
Reduced	Decreased profitability compared to	Loss of ecosystem services	
Competitiveness	pest-resistant crops (Matson et al., 1997)	(Costanza <i>et al.</i> , 1997)	
Increased Input	Investment in insecticides, pesticides	Pollution from chemical inputs	
Costs	and labor (Pretty et al., 2003)	(Pretty et al., 2005)	
Quality Degradation	Reduction in cotton fiber quality (Brevik et al., 2015)	Soil and water contamination (Scherr et al., 2008)	
Market Access	Trade restrictions due to pest-infested crops	Disruption of natural pollination processes	
Issues	(Gallai <i>et al.</i> , 2009)	(Kremen <i>et al.</i> , 2007)	
Financial Risks	Potential crop failure leading to financial	Disruption of ecosystem balance	
	losses (Tilman et al., 2002)	(Foley et al., 2005)	
Infrastructure	Damage to cotton processing facilities	Habitat destruction for other species	
Damage	(Torres et al., 2014)	(Primack et al., 1998)	

programs focus on identifying genetic traits that disrupt various stages of the bollworm's development. Researchers initiate this process by selecting cotton germplasms that display inherent resistance traits through controlled infestation trials. These germplasms are then employed in crossbreeding techniques to introduce resistance traits into commercial cotton varieties. Rigorous selection processes, incorporating phenotypic assessments and genetic analyses are essential to ensure optimal resistance levels and superior agronomic performance. Continuous testing and refinement of these new cultivars are undertaken to enhance their stability and adaptability across diverse environments. By systematically integrating resistance traits into breeding programs, researchers aim to develop cotton varieties resilient to pink bollworm infestations. This approach not only mitigates immediate pest pressures but also contributes to the long-term sustainability of cotton agriculture. Ensuring high yields and quality fiber production in the face of evolving pest challenges, these meticulous breeding strategies bolster cotton's resilience, supporting sustainable agricultural practices and securing the economic viability of cotton farming (Fig. 1).

Breeding Strategies to enhance Resistance Traits

Breeding programs aimed at enhancing cotton

resilience against pests, such as the pink bollworm (Pectinophora gossypiella), utilize a variety of techniques to augment resistance characteristics within the crop. Traditional breeding methods remain foundational, focusing on selecting and crossing cotton germplasms that exhibit existing resistance traits (Brar et al., 2018). Techniques such as recurrent selection, backcrossing, and pedigree breeding are employed to stabilize resistance genes within superior cotton cultivars while preserving desirable agronomic traits. The effectiveness of conventional breeding in developing

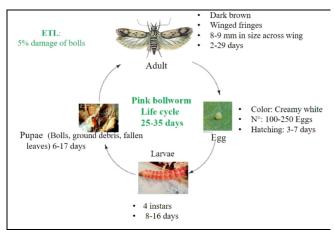


Fig. 1: Exploring the Life Cycle of the cotton Pink bollworm.

cotton varieties with heightened resistance is underscored by the utilization of genetic diversity found in both wild and cultivated cotton species (Paterson *et al.*, 2012).

Marker-assisted selection (MAS) has emerged as a pivotal molecular breeding tool in cotton agriculture, offering a streamlined approach to identifying and tracking DNA markers associated with resistance genes. Through genotyping assays, breeders can swiftly identify plants carrying favorable alleles linked to pest resistance, thereby bypassing the need for time-consuming field evaluations. This targeted approach enhances the precision and speed of cotton breeding programs, allowing breeders to focus their efforts on individuals with the highest potential for resistance (Lacape et al., 2016). Moreover, MAS enables breeders to overcome the limitations of traditional breeding methods by facilitating the incorporation of resistance genes from diverse genetic backgrounds. By identifying and tracking specific DNA markers associated with resistance traits, MAS empowers breeders to introgress multiple resistance loci into elite cultivars, thereby enhancing the durability and efficacy of pest resistance in cotton crops (Udall et al., 2006).

Recent advancements in genome editing technologies, notably CRISPR-Cas9 have revolutionized cotton breeding by providing precise methods for modifying the cotton genome to enhance resistance against pests such as the pink bollworm (Zhang et al., 2018). By leveraging CRISPR-Cas9, researchers can precisely target and edit specific genes associated with pest susceptibility, thereby conferring durable resistance without introducing foreign DNA. This technology holds significant promise for expediting the development of resistant cotton varieties by enabling precise genetic modifications tailored to combat specific pest threats (Janga et al., 2020). In contrast, transgenic methods involve the introduction of foreign genes, such as Bacillus thuringiensis (Bt) insecticidal proteins, into cotton plants to confer resistance against specific pests like the pink bollworm. The commercialization of Bt cotton cultivars has witnessed widespread adoption among farmers due to their effectiveness in controlling pest populations. However, continuous research is imperative to develop new transgenic features and deployment techniques to mitigate the emergence of resistance in insect populations (Carriere et al., 2016; Tabashnik et al., 2013).

Leveraging Genetic Engineering: Engineering Cotton for Natural Defense mechanisms

Introducing natural defense genes into cotton is a promising approach to enhance crop resilience and reduce reliance on chemical pesticides. This strategy involves identifying genes associated with natural defense mechanisms from various sources, such as wild cotton relatives, other plant species renowned for pest or disease resistance, and beneficial microorganisms. These genes encode proteins like insecticidal toxins, antimicrobial peptides, or enzymes involved in defense pathways, providing avenues to bolster cotton plants' innate ability to fend off pests and diseases (Li et al., 2013). Advanced genetic engineering techniques such as Agrobacteriummediated transformation and gene gun bombardment are employed to incorporate these defense genes into the genome. In Agrobacterium-mediated transformation, the target gene is inserted into a plasmid vector within Agrobacterium tumefaciens, a soil bacterium, which then transfers the gene into cotton plant cells. Alternatively, gene gun bombardment involves physically bombarding cotton tissues with gold or tungsten particles coated with the defense gene, facilitating its integration into the genome (Yang et al., 2013).

Following transformation, the cotton plant tissues undergo regeneration to develop into whole plants. This process involves culturing transformed cells, such as embryogenic callus or shoot apical meristem, in a growth medium supplemented with selective agents like antibiotics or herbicides linked to the inserted gene. Only cells that have successfully integrated the defense gene exhibit resistance to the selective agents and are subsequently multiplied to generate transgenic plants (Kumar et al., 2019). These transgenic cotton plants undergo rigorous characterization and screening to assess gene expression, protein synthesis, and phenotypic effects. Molecular techniques such as polymerase chain reaction (PCR) and gene expression analysis confirm the presence and functionality of the defense gene in the transgenic plants. Furthermore, field tests and bioassays are conducted to evaluate the efficacy of these plants against target pests or diseases under real-world conditions, ensuring that transgenic cotton lines with desirable agronomic traits and effective pest or disease resistance are identified for further regulatory approval and potential commercial deployment (Saha et al., 2016). Once regulatory approval is obtained, these transgenic cotton varieties offer producers an environmentally friendly and sustainable approach to pest and disease management in cotton cultivation (Pang et al., 2016).

Genetic Modification Techniques for Pest resistance

Genetic modification techniques present promising avenues for enhancing pest resistance in cotton crops, thereby reducing reliance on chemical pesticides and

promoting environmentally sustainable agricultural practices. Transgenic approaches involve the introduction of foreign genes, often sourced from bacteria or other plants, into the cotton genome to bolster natural defenses against pests like the pink bollworm. For example, genes encoding insecticidal proteins from Bacillus thuringiensis (Bt) are commonly integrated into cotton plants using methods such as gene cannon bombardment or Agrobacterium-mediated transformation (James, 2018). These transgenic cotton varieties exhibit enhanced protection against targeted pests, contributing to more resilient and productive agricultural systems. Another innovative approach, RNA interference (RNAi), leverages double-stranded RNA molecules to silence specific target genes essential for pest survival or reproduction, offering a precise and environmentally friendly method for pest management (Tabashnik et al., 2017).

Genome editing technologies, exemplified by CRISPR-Cas9, offer unprecedented precision in targeted genetic modifications, revolutionizing the prospects for pest resistance in cotton crops. By enabling the precise alteration of specific DNA sequences associated with pest susceptibility, CRISPR-Cas9 facilitates the introduction of beneficial mutations or deletions to enhance pest resistance in cotton plants (Wang et al., 2017). Moreover, CRISPR-based approaches avoid the incorporation of extraneous DNA, mitigating the risk of accelerating the emergence of resistant pest populations. Marker-assisted selection (MAS) is another powerful molecular breeding technology that streamlines the identification and selection of cotton plants with desirable pest resistance traits. By leveraging molecular markers linked to target genes associated with pest resistance, MAS accelerates the breeding process by enabling the rapid screening of breeding populations for desired traits, thus contributing to the development of more resilient and productive cotton cultivars tailored to meet the challenges of pest pressure in agricultural landscapes (Janga et al., 2020).

Cultural Control methods

Cultural control methods are fundamental to IPM strategies in cotton agriculture, aimed at creating environments unfavorable for pest development and reproduction. Crop rotation, for instance, involves alternating cotton cultivation with non-host crops, disrupting pest life cycles and reducing their soil accumulation. Incorporating grains or legumes in rotation sequences effectively disrupts pest cycles while replenishing soil nutrients, enhancing the sustainability of

cotton farming (Lu *et al.*, 2010). Additionally, plowing techniques play a critical role in cultural control. Conservation tillage practices disrupt overwintering habitats of pests, thereby decreasing their survival rates. These methods also contribute to soil conservation by mitigating erosion and preserving soil organic matter and moisture content (Reisig *et al.*, 2018).

Biological Control agents

Biological control agents use competition, parasitism, or predation to manage pest populations in cotton cultivation. Predators such as lady beetles, lacewings, and spiders naturally prey on cotton pests like aphids, mites, and caterpillars, thus contributing to sustainable pest management (Colazza *et al.*, 2004). Parasitoids, including parasitic wasps, lay their eggs within pest insects such as caterpillars, aphids, and whiteflies. The larvae then develop by consuming the host from within, effectively reducing pest populations and safeguarding cotton crops (Desneux *et al.*, 2007). Utilizing these biological control agents enables cotton farmers to reduce their dependence on chemical pesticides, promoting environmentally friendly pest management practices conducive to long-term agricultural sustainability.

Bioinformatics and Genomics

Bioinformatics and genomics are indispensable tools in elucidating the genetic mechanisms underlying resistance in cotton against pests like the pink bollworm. bioinformatics facilitates the study and interpretation of biological data through computer-based techniques and tools. In the context of pest resistance in cotton, bioinformatics streamlines several key processes. Firstly, sequence analysis enables the identification and comparison of DNA sequences from diverse cotton cultivars or closely related species, aiding in the detection of genetic variants associated with insect resistance (Loman et al., 2013). Additionally, cotton genome annotation plays a crucial role by identifying genes, regulatory elements, and functional regions implicated in pest resistance, thus providing valuable insights into the genetic basis of resistance traits. Furthermore, data integration is pivotal in gaining a comprehensive understanding of the molecular processes governing pest resistance (Schuler, 1997). By integrating various datasets, including genomic, transcriptomic, and proteomic data, researchers can discern intricate molecular interactions and pathways involved in cotton defense mechanisms against pests. Overall, the synergy between bioinformatics and genomics holds immense promise in unraveling the genetic architecture of pest resistance in cotton, offering valuable insights for the development of

Year	Historical Context and Spread of Pink Bollworm	Organization Responsible
1917	First recorded in the United States in the southwestern states, primarily Arizonaand California.	United States Department of Agriculture (USDA)
1921	Pink bollworm identified as a major pest in cotton production, causing significant crop losses.	USDA
1950s	Pink bollworm spreads to cotton-growing regions across the southern United States.	USDA
1970s	Pest establishes itself in Texas and spreads further into other cotton-growing states.	USDA, National Cotton Council
1980s	Pink bollworm becomes a widespread pest in cotton fields throughout the southern and western United States.	USDA, National Cotton Council
1990s	Efforts to control pink bollworm intensify with the implementation of integrated pest management strategies.	USDA, National Cotton Council, State Agricultural Departments
2000s	Introduction of genetically modified (GM) cotton varieties engineered for pink bollworm resistance.	USDA, Biotechnology Companies
Present	Pink bollworm continues to pose challenges to cotton agriculture, albeit with reduced impact due to control measures and resistant cotton varieties.	USDA, Agricultural Research Institutions

Table 2: The name of the organization responsible for establishing the historical context and spread of the pink bollworm.

resilient cotton cultivars.

Genome Sequencing and Annotation of cotton

Genome sequencing and annotation have revolutionized our ability to unravel the genetic basis of cotton pest resistance. With the advent of high-throughput sequencing technologies, comprehensive genome assemblies for key cotton species such as Gossypium hirsutum and Gossypium barbadense have been realized. One of the primary utilities of these genome sequences lies in gene discovery. Through comparative genomics and association studies, researchers can identify candidate genes and genomic regions associated with pest resistance. By analyzing sequence variations among different cotton cultivars or related species, researchers can pinpoint genetic loci that play crucial roles in conferring resistance to pests (Li et al., 2014). Moreover, functional annotation of the cotton genome enables researchers to decipher the functions of individual genes and regulatory elements implicated in pest resistance pathways. This annotation process provides insights into the molecular mechanisms underlying cotton defense mechanisms against pests.

Scientific advancements with agricultural practices

Integrating scientific discoveries with agricultural practices is crucial for fostering sustainable farming among cotton farmers. Extension services play a pivotal role in facilitating this integration by disseminating pertinent scientific knowledge and best practices through seminars, field demonstrations and outreach initiatives.

These programs, often led by agricultural specialists and researchers, not only impart valuable insights into sustainable crop production and pest management but also foster effective communication between scientists and farmers, ensuring the translation of scientific knowledge into practical solutions tailored to regional agricultural contexts (Table 2).

Moreover, educational programs and training sessions are indispensable for equipping farmers with the necessary skills and knowledge to implement sustainable farming methods successfully. Workshops, seminars and online courses provide farmers with valuable information on soil health management, water conservation techniques, integrated pest management (IPM) strategies, and other sustainable agricultural practices (FAO, 2015). By empowering farmers with practical skills and scientific understanding, these educational initiatives enable them to make well-informed decisions and adapt to evolving agricultural challenges while minimizing environmental impact.

Furthermore, fostering farmer adoption of sustainable agricultural techniques necessitates evaluating their long-term sustainability and economic feasibility. Economic analysis tools, such as cost-benefit evaluations and return on investment estimates, enable farmers to compare the financial implications of implementing sustainable practices against those of traditional methods (Tegtmeier *et al.*, 2005). Long-term sustainability assessments consider variables such as climate change resilience, water

usage efficiency, biodiversity conservation, and soil health. By demonstrating the economic viability and benefits of sustainable farming, these evaluations support the adoption of resilient and ecologically friendly agricultural systems.

The future scope

The future of cotton breeding for sustainable pest management relies heavily on the integration of emerging technologies, offering innovative solutions to combat pests while minimizing environmental impact. Key technologies like precision agriculture, remote sensing, and unmanned aerial vehicles (UAVs) have the potential to revolutionize pest management practices. They enable targeted application of inputs, early detection of pest outbreaks, and real-time monitoring of crop health. Notably, utilizing UAVs for the precise delivery of biopesticides or pheromones has demonstrated significant reductions in pesticide usage and non-target effects, thereby advancing sustainable pest management (Mulla *et al.*, 2020).

Nevertheless, the challenge of controlling the emergence of pest resistance remains a significant concern, necessitating ongoing innovation and proactive management techniques. The development of resistant pest populations often stems from an over-reliance on singular pest control methods, such as chemical pesticides or genetically engineered crops (Tabashnik *et al.*, 2013). Integrated pest management (IPM) strategies, which incorporate diverse control techniques including chemical, biological, and cultural treatments, are crucial for mitigating resistance. Additionally, the progression of novel pest management technologies such as gene editing and RNA interference (RNAi) holds promise for overcoming resistance and bolstering the sustainability of pest control methods (Rajeswaran *et al.*, 2016).

Conclusion

In conclusion, the future of sustainable pest management in cotton agriculture hinges on a multifaceted approach that integrates innovative technologies, proactive resistance management strategies and careful consideration of socio-economic implications. By harnessing emerging technologies such as precision agriculture and UAVs, growers can optimize pest control measures, while minimizing environmental impact. Addressing resistance development in pests requires the adoption of integrated pest management (IPM) strategies that combine cultural, biological and chemical control methods, alongside the development of novel pest control technologies like RNA interference and gene editing. Moreover, assessing the socio-economic implications of pest management strategies is crucial for ensuring their feasibility and adoption by farmers. Through collaboration

between researchers, stakeholders and policymakers, we can overcome challenges and promote the development of sustainable pest management practices that safeguard the long-term health and productivity of cotton crops, while supporting the livelihoods of cotton growers and contributing to a more sustainable agricultural future.

Conflict of interest

No authors declared conflict of interest.

References

- Altieri, M.A. and Nicholls C.I. (2003). Soil fertility management and insect pests, harmonizing soil and plant health in agroecosystems. *Soil Till. Res.*, **72(2)**, 203-211.
- Brar, G.S. and Singh K. (2018). Breeding cotton for resistance to insect pests: Challenges and achievements. *SABRAO J. Breed. Genet.*, **50**(1), 1–24.
- Brar, M.S., Gill H.S. and Chahal K.K. (2018). Evaluation of Cotton Germplasm for Resistance to Pink Bollworm. *J. Cotton Res.*, **12**(3), 45-53.
- Brevik, E.C., Sullivan P., Parikh S.J., Pillai-McGarry U. and Steiner J.L. (2015). The potential role of cotton (*Gossypium hirsutum* L.) in reducing erosion and improving soil health in the southeastern United States. *J. Soil Water Conserv.*, **70(2)**, 46A-50A.
- Carriere, Y., Crickmore N. and Tabashnik B.E. (2016). Optimizing pyramided transgenic Bt crops for sustainable pest management. *Nat. Biotechnol.*, **34(3)**, 289–292.
- Chapin Iii, F.S., Zavaleta E.S., Eviner V.T., Naylor R.L., Vitousek P.M., Reynolds H.L. and Díaz S. (2000). Consequences of changing biodiversity. *Nature*, **405**(**6783**), 234-242.
- Colazza, S., Fucarino A., Peri E., Salerno G, Conti E. and Bin F. (2004). Insect oviposition induces volatile emission in herbaceous plants that attracts egg parasitoids. *J. Exp. Biol.*, **207(1)**, 47-53.
- Costanza, R., d'Arge R., De Groot R., Farber S., Grasso M., Hannon B. and Van Den Belt M. (1997). The value of the world's ecosystem services and natural capital. *Nature*, **387(6630)**, 253-260.
- Dennehy, T.J. (2000). Effects of plant resistance and parasitism on the population dynamics of the pink bollworm (Lepidoptera, *Gelechiidae*). *Environ. Entomol.*, **29(2)**, 341–346
- Desneux, N., Decourtye A. and Delpuech J.M. (2007). The Sublethal Effects of Pesticides on Beneficial Arthropods. *Annu. Rev. Entomol.*, **52(1)**, 81–106.
- Dhillon, M.K., Sharma H.C. and Arora R. (2021). Integrating host plant resistance and biopesticides for sustainable pest management. *J. Pest Sci.*, **94(1)**, 225-243.
- FAO (2015). Sustainable agricultural development for food security and nutrition, What roles for livestock? Food and Agriculture Organization of the United Nations.
- Foley, J.A., DeFries R., Asner G.P., Barford C., Bonan G., Carpenter S.R. and Snyder P.K. (2005). Global consequences of land use. *Science*, **309**(**5734**), 570-574.

- Gallai, N., Salles J.M., Settele J. and Vaissière B.E. (2009). Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. *Eco. Econ.*, **68(3)**, 810-821.
- Godfray, H.C.J., Beddington J.R., Crute I.R., Haddad L., Lawrence D., Muir J.F. and Toulmin C. (2010). Food security, the challenge of feeding 9 billion people. *Science*, **327**(**5967**), 812-818.
- Gore, J., Catchot A.L. and Cook D.R. (2022). Principles and practices of integrated pest management for cotton. *Annu. Rev. Entomol.*, **67**, 23-42.
- Gurr, G.M., Lu Z., Zheng X., Xu H., Zhu P., Chen G. and Heong K.L. (2016). Multi-country evidence that crop diversification promotes ecological intensification of agriculture. *Nat. Plants*, **2(3)**, 1-4.
- James, C. (2018). Global Status of Commercialized Biotech/ GM Crops in 2018, Biotech Crop Adoption Surges as Economic Benefits Accumulate in 23 Years. ISAAA Brief No. 54.
- Janga, M.R., Campbell L.M. and Rathore K.S. (2020). CRISPR/Cas9-mediated targeted mutagenesis in *Cotton* (*Gossypium hirsutum* L.). *Plant Mole. Biol. Reporter*, **38(1)**, 1–9.
- Kremen, C., Williams N.M., Aizen M.A., Gemmill Herren B., LeBuhn G., Minckley R. and Ricketts T.H. (2007). Pollination and other ecosystem services produced by mobile organisms, a conceptual framework for the effects of land use change. *Eco. Lett.*, **10(4)**, 299-314.
- Kumar, S. and Singh N.K. (2019). Transgenic cotton, current status, benefits and limitations. *Appl. Biol. Res.*, **21**(2), 123-128.
- Lacape, J.M., Jacobs J., Arioli T., Derijcker R., Forestier-Chiron N., Llewellyn D. and Giband M. (2016). Meta-analysis of cotton fiber quality QTLs across diverse environments in a *Gossypium hirsutum* × *G. barbadense* RIL population. *BMC Plant Biol.*, **16(1)**, 88.
- Lal, R. (2009). Soil degradation as a reason for inadequate human nutrition. *J. Sustain. Agric.*, **29(2)**, 3-23.
- Li, F., Fan G, Wang K., Sun F., Yuan Y., Song G and Wang J. (2014). Genome Sequence of the Cultivated Cotton (*Gossypium arboreum*). *Nat. Genetics*, **46(6)**, 567–572.
- Li, S., Zhang X., Wang W., Guo W. and Zhang T. (2013). Inheritance and Fine Mapping of Si 1, a Silicon Accumulation QTL on the Short Arm of Chromosome 3 in Upland Cotton. *PLoS ONE*, **8**(7), e67740.
- Liu, Q., Chen Y., Gao Y. and Hou Y. (2017). Advances in breeding of new cotton varieties resistant to pests. *Mol. Plant Breed.*, **15**(7), 2385–2391.
- Loman, N.J. and Watson M. (2013). Successful Test Launch for Nanopore Sequencing. *Nat. Methods*, **10(5)**, 451–454.
- Lu, Y., Wu K., Jiang Y., Xia B., Li P., Feng H. and Cui J. (2010). Mirid Bug Outbreaks in Multiple Crops correlated with Wide-scale Adoption of Bt cotton in China. *Science*, 328(5982), 1151–1154.

- Matson, P.A., Parton W.J., Power A.G. and Swift M.J. (1997). Agricultural intensification and ecosystem properties. *Science*, **277**(**5325**), 504-509.
- Mulla, D.J., Klamke M.A. and Naranjo S.E. (2020). Drones for Precision Agriculture, Opportunities and challenges. *J. Econ Entomol.*, **113(1)**, 1–8.
- Pang, Y., Zhu Y. and Xie X. (2016). Cotton Proteomics for Discovering New Proteins in Genetically Modified (GM) Crops. In: Adv. Agron., (Vol. 139, pp. 157-191). Academic Press.
- Paterson, A.H., Wendel J.F., Gundlach H., Guo H., Jenkins J., Jin D. and Hafez A.N. (2012). Repeated polyploidization of Gossypium genomes and the evolution of spinnable cotton fibres. *Nature*, **492(7429)**, 423–427.
- Peng, X., Sun D., Zhao Y. and Li Z. (2020). Genomic insights into cotton adaptation and evolution. *J. Cotton Res.*, **3(1)**, 3.
- Pimentel, D., Wilson C., McCullum C., Huang R., Dwen P., Flack J. and Cliff B. (1997). Economic and environmental benefits of biodiversity. *BioScience*, **47**(11), 747-757.
- Pretty, J. (2007). Agricultural sustainability, concepts, principles and evidence. *Philosophical Trans. Royal Soc. B: Biol. Sci.*, **363(1491)**, 447-465.
- Pretty, J.N., Brett C., Gee D., Hine R.E., Mason C.F., Morison J.I. and van der Bijl G. (2000). An assessment of the total external costs of UK agriculture. *Agric. Systems*, **65(2)**, 113-136.
- Pretty, J.N., Noble A.D., Bossio D., Dixon J., Hine R.E., Penning de Vries F.W. and Morison J.I. (2006). Resource-conserving agriculture increases yields in developing countries. *Environ. Sci. Technol*.
- Primack, R.B. (1987). Relationships among flowers, fruits and seeds. *Annu. Rev. Ecol. Syst.*, **18(1)**, 409-430.
- Rajeswaran, J. and Pooggin M.M. (2016). RNA Interference, Relevance and Development in Agricultural Research. *In vitro Cell. Develop. Biol. Plant*, **52(2)**, 135–147.
- Razzaq, A., Sadia B., Ali S., Qayyum A. and Khan S.N. (2019). Omics approaches for improving cotton resistance to pests. In: *Omics Approaches for Sugarcane Crop Improvement* (pp. 215-233).
- Reisig, D.D. (2018). Cultural Control Tactics for Managing Insect Pests of Row Crops. *Insects*, **9(4)**, 168.
- Saha, S., Sarker A. and Adak M.K. (2016). Genetic Engineering for Enhancement of Plant Productivity. *Crop Improv. Microb. Biotechnol.* (pp. 331-347). Academic Press.
- Scherr, S.J. and McNeely J.A. (2008). Biodiversity conservation and agricultural sustainability, towards a new paradigm of 'ecoagriculture' landscapes. *Philosophical Trans. Royal Soc.: Biolog. Sci.*, **363(1491)**, 477-494.
- Schuler, G.D. (1997). Pieces of the Puzzle, Expressed Sequence Tags and the Catalog of Human Genes. *J. Mol. Med.*, **75(10)**, 694–698.
- Sharma, R., Jhala R.C. and Puri S.N. (2022). Environmental consequences of pesticide use in agriculture. *Environ. Monit. Assess.*, **194(2)**, 115.

Smith, C.M. and Reynolds O.L. (2019). Insect resistance management, Biology, economics and prediction. Academic Press.

- Smith, J.L., Jones K. and Wright D. (2012). Backcrossing and Line Development for Pink Bollworm Resistance in Cotton. *Plant Breed. Rev.*, **36**, 45-78.
- Tabashnik, B.E. and Carriere Y. (2017). Surge in insect resistance to transgenic crops and prospects for sustainability. *Nat. Biotechnol.*, **35(10)**, 926–935.
- Tabashnik, B.E., Brevault T. and Carriere Y. (2013). Insect resistance to Bt crops, Lessons from the first billion acres. *Nat Biotechnol.*, **31(6)**, 510–521.
- Tegtmeier, E.M. and Duffy M.D. (2005). External Costs of Agricultural Production in the United States. *Int. J. Agric. Sustain.*, **3(1)**, 1–20.
- Tilman, D., Cassman K.G., Matson P.A., Naylor R. and Polasky S. (2002). Agricultural sustainability and intensive production practices. *Nature*, **418**(**6898**), 671-677.
- Tilman, D., Reich P.B. and Knops J.M. (2006). Biodiversity and ecosystem stability in a decade-long grassland experiment. *Nature*, **441(7093)**, 629-632.
- Torres, A.B., Costa H.C., Queiroz H.L., Scariot A. and Lopes A.V. (2014). A spatial approach to assessing habitat loss and fragmentation due to disturbance by cotton agriculture in the Brazilian *Cerrado. J. Environ. Manage.*,

- **145**, 277-284.
- Tylianakis, J.M., Didham R.K., Bascompte J. and Wardle D.A. (2008). Global change and species interactions in terrestrial ecosystems. *Ecol Lett.*, **11(12)**, 1351-1363.
- Udall, J.A. and Wendel J.F. (2006). Polyploidy and crop improvement. In: *Plant Breed. Rev.* (Vol. **27**, pp. 35–57). John Wiley & Sons, Inc.
- Vaughan, M.M., Huffaker A., Schmelz E.A., Dafoe N.J., Christensen S.A. and Sims J. (2018). Integrating multiomics for agricultural development. *Front. Plant Sci.*, **9**, 121.
- Vennila, S., Deshmukh A.J. and Solanki H.G. (2021). Impact of pink bollworm on cotton production. *Cotton Res. J.*, **12(3)**, 221-230.
- Wang, X. and Ma Y. (2017). Genetic Engineering of Cotton with Insecticidal Traits for Pest Resistance. In: *Cotton* (pp. 79–100). Springer, Cham.
- Yang, X., Li X., Zhang X., Gao S., Wu M. and Xia Z. (2013). Overexpression of Cotton Gene GhPFN2 enhances Disease Resistance in Transgenic Tobacco. *PLoS ONE*, **8(12)**, e 66868.
- Zhang, B., Yang X., Yang C., Li M. and Guo Y. (2018). Exploiting the CRISPR/Cas9 system for targeted genome mutagenesis in *Petunia*. *Sci. Hortic.*, **240**, 577–584.